

MARINONE AND DEBROMOMARINONE: ANTIBIOTIC SESQUITERPENOID NAPHTHOQUINONES OF A NEW STRUCTURE CLASS FROM A MARINE BACTERIUM

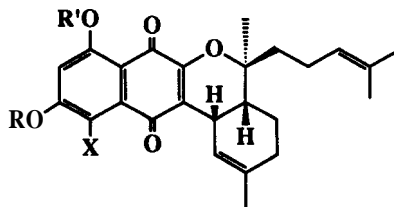
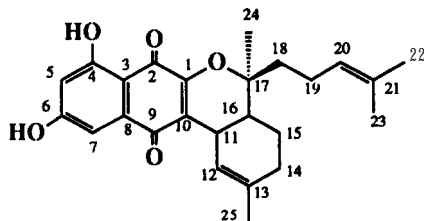
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Summary - Marinone (1) and its debromo analog debromomarinone (2), antibiotic sesquiterpenoid naphthoquinones of a new structure class, have been isolated from the organic extract of the liquid culture of a marine actinomycete, isolate CNB-632. The structures of the new compounds were assigned on the basis of comprehensive spectroscopic analyses.

Although the production of antibiotic compounds by terrestrial bacteria has been the focus of intense investigation for over 50 years, analogous investigations of marine bacteria have been severely limited.¹ Due to problems in the isolation and identification of marine bacteria and fungi, the immensity of this resource has yet to be demonstrated. In preliminary chemical studies of several classes of marine bacteria, a variety of interesting new molecules have recently been discovered.^{1,2} In this paper, we report the isolation and structural elucidation of marinone (1), a sesquiterpenoid naphthoquinone and its debromo analogue, debromomarinone (2), new **antibacterials** of a novel class, isolated from the culture broth of an unidentified marine actinomycete. The new molecules are among a rare group of bacterial metabolites which are produced via mixed biosynthesis involving both acetate and terpene pathways.

The microorganism, culture # CNB-632,³ was grown in 10L culture and the whole broth was repetitively extracted with ethyl acetate. Concentration of the extract under vacuum yielded 1.73 g of crude material, which was



- 1 : R = R' = H, X = Br
2 : R = R' = X = H
3 : R = Me, R' = X = H

fractionated by flash chromatography on silica using a gradient of ethyl acetate in isoctane. Fractions eluted with 30% ethyl acetate / isoctane were combined and further purified by normal phase HPLC and reversed-phase preparative TLC (70% MeOH/H₂O) to obtain 14 and 2,⁵ each as red, non-crystalline solids.

The mass spectrum of 1 showed a doublet of equal intensity at *m/z* 486 and 488, which by HR mass analysis clearly indicated a molecular formula C₂₅H₂₇BrO₅ containing bromine. Marinone (1) and 2 possessed very similar ¹H NMR, IR and UV spectra. In the mass spectrum of 2, a parent ion was observed at *m/z* 408 which implied that 2 was the debromo analog of 1. This was confirmed when the molecular formula analysed for C₂₅H₂₈O₅ by HREIMS. The proton NMR spectra of 1 and 2 were simple and relatively uninformative. However, analysis of the ¹H NMR COSY spectrum of 1 led to the assignment of several structural fragments. A resonance at δ 5.08 (bt, J = 7 Hz) coupled to a signal at δ 2.0 and to two methyl resonances at δ 1.49 and 1.66. in combination with mass spectral loss of an *m/z* = 83 fragment to give a strong peak at *m/z* 325. indicated a C₆H₁₁ terpenoid side-chain in the molecule. Proton NMR COSY correlations involving three methine protons at δ 1.88, 3.39 and 6.03 identified a fragment, -CH₂CHCHCH=CCH₃-, assigned to C15, C16, C1, and C12. These two fragments were linked through a quaternary carbon (C17) using long range proton-carbon correlations acquired through HMBC experiments. A two bond correlation from a methyl resonance (C24) at δ 1.49 to C17 at δ 83.5, and a three bond correlation to C18 at δ 37.5 linked C24 and C18 through C17. Another three bond correlation from the same methyl protons to C16 linked

Table. NMR Data for Debromomarinone (2)*

C#	Carbon NMR Data (ppm)	Proton NMR Data (δ)	HMBC Data
1			
2	155.1 (C)		H11
	180.9 (C)		
3	105.6 (C)		H5, H7
			H5
4	107.9 (CH)	6.10 (d, 2.5)	H7
6	167.2 (C)a		H5
7	115.6 (CH)	6.83 (d, 2.5)	H5
8	135.7 (C)		
9	186.4 (C)		H7
10	123.5 (C)		H11, H16
12	32.0 (CH)	3.39 (bt, 5.0)	H12, H16
13	121.7 (CH)	6.03 (bd, 5.0)	H11, H14, H25
14	130.7 (C)		H11, H14, H25
	(CH2)	2.00 (m)	H12, H25
15	21.4 (CH2)	1.31 (dt, 6.0, 12.0, 12.0)	H11
		2.00 (m)	
17	39.1 (CH)	1.88 (ddd, 3.0, 6.0, 12.0)	H11, H12, H14, H24
18	37.5 (CH2)		H24
		1.63 (m)	H19, H24
19	23.2 (CH2)	2.00 (m)	H20
20	124.8 (CH)	5.08 (bt, 7.0)	H19, H22, H23
21	132.9 (C)		H19, H22, H23
23	25.8 (CH3)	1.60 (bs)	H20
24	22.7 17.6 (CH3)	1.51 1.49 (bs)	H20
	(CH3)	(s)	
25	23.8 (CH3)	1.66 (bs)	H12

* Spectra were acquired in CD₃OD with reference to internal TMS at 0.00 ppm for ¹H (500 MHz) and CD₃OD at 49.0 ppm for ¹³C (50 MHz). Assignments are based on the results of XHCORR (50 MHz) and HMBC (500 MHz) experiments. a- assignments may be interchanged.

C16 and C17. Long range correlations from the C14 methylene protons to C12, C13 and C16 allowed the terpenoid portion of the molecule to be fully defined as in **1** and **2**.

The remaining aromatic portion of the molecule accounted for 9 degrees of unsaturation, and was characterized by two ^1H NMR resonances at δ 6.83 and 6.10 ppm and ^{13}C NMR resonances for eight **sp² quaternary** carbons, including two quinone carbonyl resonances at δ 186.4 and 180.9. This information, combined with the UV-Vis absorption characteristics of these compounds, **4, 5** (absorbances between 480-510 nm) and the **IR** absorption at 1638 cm^{-1} of **1** suggested a hydroxynaphthoquinone moiety. **6, 7** In the ^1H NMR spectrum of **2**, the 2.5 Hz coupling between the two aromatic protons at δ 6.10 and 6.83 indicated that they were *meta* oriented, thus allowing their assignment at **C5** and **C7**, respectively. The presence of two phenolic OH groups was confirmed by the ^1H NMR characteristics of **3**, the monomethyl ether derivative produced by the treatment of **2** with **diazomethane**. **8** Because of intense hydrogen bonding with the **C9** carbonyl, the hydroxyl at **C7** was not methylated under the reaction conditions. The hydroxyl proton appeared at δ 12.0 ppm in the ^1H NMR spectrum of **3**, a shift common in this hydrogen bonded arrangement.* The protons of the methyl ether showed strong **nOe** when either of the aromatic protons were irradiated, allowing the methyl ether to be established at C6.

The naphthoquinone component of debromomarinone was linked to the terpenoid moiety using HMBC correlations. This included a **3-bond** correlation from **H1** to C1 and two and three bond correlations from **H11** and **H16** to **C10**, respectively. To satisfy the molecular formula, C1 had to be linked to C17 via a chromanol ether linkage. Finally, the substitution pattern on the aromatic ring was successfully established using HMBC results in combination with ^{13}C NMR data. In naphthoquinone chromenols with **peri-substituted** hydroxyl groups, the carbonyl adjacent to the chromenol oxygen-bearing carbon is known to experience a significant **upfield** shift.⁹ On this basis, the carbonyl resonances at δ 180.9 and 186.4 were assigned to **C2** and **C9**, respectively. A strong **3-bond** correlation from H7 to **C9** in an HMBC experiment with **2** enabled H7 to be assigned, thus providing the regiochemistry of the aromatic ring. The ^1H NMR spectra of **1** and **2** were almost superimposable except for the **aromatic** proton signal assigned at C7 in **2**. Since this signal was absent in marinone (**1**), the bromine substituent was placed at this position.

The relative configurations at the three **chiral** centers in **1** and **2** were determined using ^1H NMR **nOe** results. A strong **nOe** in **2** from H1 to H16, and vice versa, indicated a **cis** configuration of the C1-C16 ring junction. The **C17-C18** bond was assigned as pseudoaxial (β) on the basis of an **nOe** between H1 and one of the protons at C18. Curiously, both marinone and **debromomarinone** displayed no optical rotation at the sodium D line, indicating that they may be racemic.

Marinone and its debromo derivative are the first examples of compounds possessing this novel carbon skeleton. Two related compounds, **napterpin**,¹⁰ and naphthogeranine **A**,¹¹ both isolated recently from the fermentation broths of terrestrial actinomycetes, possess similar chromenol C and D rings, constructed in these cases from monoterpenoid substituents. Marinone and debromomarinone are formally the sesquiterpenoid analogs of these latter compounds. Both metabolites show significant in vitro antibacterial activity mainly against gram-positive bacteria. For example, **marinone** (**1**) shows MIC = 1 $\mu\text{g/ml}$ against *Bacillus subtilis*, and debromomarinone shows MIC values of 1-2 $\mu\text{g/ml}$ against *Staphylococcus aureus*, *S. epidermis*, *S. pneumoniae*, *S. pyogenes* and *S. epidermis*.

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3. The bacterium, isolate CNB-632. was obtained by serial dilution and plating techniques, using media containing rifampicin (5 µg/ml), from a sediment sample collected at the Torrey Pines Estuary, La Jolla, CA in 1989. The isolate was cultured in a marine medium (A1) consisting of 1% starch, 0.4% yeast extract, 0.2% peptone, 75% seawater, 25% deionized water, buffered with 10 ml l .O M Tris buffer and shaken at 230 RPM for 7 days at 23°C. The whole culture was extracted with EtOAc to leave an extract from which compounds 1 (2-5 mg/L) and 2 (7-10 mg/L) were isolated. Isolate CNB-632 is an actinomycete which shows brown vegetative mycelia and lacks aerial spores and hyphae (A1 medium).
4. For Marinone (1): UV-Vis (MeOH) 508 (ε 4100), 406 (ε 2290), 300 (ε 10500). 235 (ε 17400) nm; IR (film) 3417 (br), 2919, 1638, 1585, 1545, 1452, 1393, 1323, 1244 cm⁻¹; LREIMS *m/z* 488 (20), 486 (13), 408 (34), 325 (65), 323 (86), 28 (100) ; HREIMS *m/z* 486.0996. calcd. for C₂₅H₂₇BrO₅ 486.1039; ¹H NMR (200 MHz, CD₃OD) δ 6.10 (s, 1H), 5.95 (d, J = 5 Hz, 1H), 5.03 (bt, J = 7 Hz, 1H), 3.40 (br, 1H), 2.0 (m, 5H), 1.66 (bs, 3H), 1.60 (bs, 3H), 1.5 (m, 2H), 1.55 (bs, 3H), 1.45 (s, 3H).
5. For Debromomarinone (2): UV-Vis (MeOH) 488 (ε 2280). 388 (ε 2600), 299 (ε 9200), 269 (ε 10400), 226 (ε 16600) nm; IR (film) 3414(br), 2924, 1631, 1589, 1544, 1469, 1380, 1228 cm⁻¹; LRBIMS *m/z* 408 (M⁺, 60), 325 (100), 297 (30); LRCIMS *m/z* 409 (M⁺+ H); HREIMS *m/z* 408.1916, calcd. for C₂₅H₂₈O₅ 408.1924, ¹H and ¹³C NMR: see table.
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8. For 3 : IR (film) 2926, 1638, 1591, 1446, 1388, 1312, 1224, 1160 cm⁻¹; ¹H NMR (500 MHz, CDC13) δ 12.0 (s, 1H), 7.15 (d, J = 2.5 Hz, 1H), 6.55 (d, J = 2.5 Hz, 1H), 6.07 (d, J = 5.5 Hz, 1H), 5.02 (bt, J = 5.5 Hz, 1H), 3.46 (bt, J = 5 Hz, 1H), 2.0 (m, 5H), 1.88 (ddd, J = 3, 6.5, 12 Hz, 1H), 1.68 (bs, 3H), 1.65 (m, 2H), 1.62 (bs, 3H), 1.56 (bs, 3H), 1.54 (s, 3H), 1.34 (m, 1H).
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